

Guidelines for transoesophageal echocardiographic probe cleaning and disinfection from the British Society of Echocardiography[†],[‡]

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The clinical utility of transoesophageal echocardiography (TOE) is well established. Being a semi-invasive procedure, however, the potential for transmission of infection between sequential patients exists. This has implications for the protection of both patients and medical staff. Guidelines for disinfection during gastrointestinal endoscopy (GIE) have been in place for many years. ^{1,2} Unfortunately, similar guidance is lacking with respect to TOE. Although traversing the same body cavities and sharing many similarities with upper GIE, there are fundamental structural and procedural differences with TOE which merit special consideration in establishing a decontamination protocol. This document provides recommendations for TOE probe decontamination based on the available evidence, expert opinion, and modification of the current British Society of Gastroenterology guidelines.

Keywords

Probe cleaning • Probe disinfection • Transoesophageal echocardiography

Introduction

The basic principles underpinning successful decontamination of reusable equipment are cleaning and either manual or automated disinfection. Transoesophageal echocardiographic (TOE) probes do not warrant sterilization, as they are endoscopes not penetrating sterile areas of the body (unlike laparoscopes or other surgical instruments), nor is sterilization a feasible option.

Structural design of a transoesophageal echocardiographic probe

All TOE probes, irrespective of the manufacturer, share the same components (see Glossary of terms; *Figure 1*): a flexible tip, a shaft, a probe handle with steering controls, and a cable which attaches to the plug socket in its associated equipment (*Figure 1*). Handling of any part of this apparatus results in a potential source of infection (*Figure 2*; these were probes considered to have been decontaminated by a manual wipe system and were ready for re-use with a subsequent patient). During passage into a patient, there is

contact with intact mucous membranes and the potential for contact with non-intact mucous membranes. This means that TOE probes require disinfection to a similar level as upper gastro-intestinal endoscopy (GIE).³

A TOE probe is flexible, reusable, delicate, expensive, and heatsensitive. A TOE probe cannot withstand the standard techniques of sterilization utilizing heat and steam. Although gas sterilization is possible, the high costs and long cycle times involved at present render this technique impractical for routine use.

Advantages over gastrointestinal endoscopy

Unlike standard upper GIE, TOE probes do not have internal channels for air, water, or biopsy, which reduces exposure to those risks that prompted greater awareness of UK decontamination practices in 2004 (MHRA Medical Device Alert 2004/028)³ and makes contamination far more accessible for removal and disinfection.

Disadvantages over gastrointestinal endoscopy

The probe handle, including the steering mechanism and the plug socket, is not sealed and cannot be immersed in any liquid for

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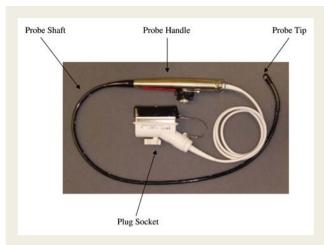


Figure 1 Components of a standard transoesophageal echocardiographic probe.



Figure 2 Any part of the transoesophageal echocardiographic probe can be a source of contamination (Photograph: Peter Hoffman).

cleaning or disinfection. Entry of fluid or contamination in this region may result in corrosion and damage to electrical connections as well as serve as a vector for infection.⁴

Transmission of infection during transoesophageal echocardiography

Evidence that there is a risk of cross-infection is minimal, but the absence of evidence is not evidence of the absence of risk. Estimating the infection risk is difficult for several reasons. First, there are

Table I Examples of agents potentially transmissible by transoesophageal echocardiography

Cross-infection from patient to patient and patient to staff

Bacteria—Helicobacter pylori, Pseudomonas aeruginosa, Salmonella species, Mycobacterium species

Viruses-Hepatitis B and C, HIV

Prions—vCJD

Contamination of patients from the decontamination procedure

Bacteria—Pseudomonas aeruginosa, Legionella pneumophila,

Mycobacterium species

no well-performed comprehensive studies relating to infection control in TOE practice. Secondly, the onset of infections relating to procedures may be delayed until after discharge from hospital making diagnosis and reporting unlikely. As a marker for comparison, the reported frequency of infection following upper GIE is 1 in 1.8 million studies.⁵ Based on this wider reporting in relation to upper GIE and bronchoscopy, it seems sensible to implicate the same causative agents during TOE⁵⁻⁷ (*Table 1*). Thirdly, the possibility remains of transmission of infectious microorganisms from one individual to another via the TOE probe with very long incubation periods. The issue of prion infection will be discussed in a later section.

There are no data relating to patient-patient risk of infection during TOE. The aim of decontamination is the prevention of cross-infection from one individual to another, which is the focus of these guidelines.

Recommendations for transoesophageal echocardiography decontamination

Effective decontamination strategies should promote health and safety of staff and patients alike. Care should be taken to safeguard TOE probe integrity and prevent damage that renders manufacturer warranty inoperable. Therefore, an ideal decontamination policy should consider the following:

- (i) Health and safety at work
- (ii) Manufacturer warranties
- (iii) Appropriate use of disinfectants
- (iv) Clear delineation of 'clean' and 'dirty' areas in TOE probe decontamination and storage
- (v) A robust technique of decontamination
- (vi) Training and provision of staff to engage in quality assurance audit and ensure traceability

General considerations

Health and safety

The Health and Safety at Work etc. Act 1974⁸ (HSAWA) requires employers to ensure the safety and welfare of all employees as far as is reasonably practicable. In turn, employees should

comply with the established precautions to safeguard their working environment. A risk assessment should be performed before selection and purchase of an appropriate disinfectant for use with TOE. There are numerous disinfectants currently available on the market which may have potential hazardous effects. Further guidance is available from The Control of Substances Hazardous to Health Regulations 1994⁹ (COSHH).

Manufacture warranty

Prior to purchase of a TOE probe, an Echo department should review the manufacturer's recommendations regarding decontamination to ensure that these can be carried out within the resources available. This is important for a number of reasons. First, adherence to such instructions is mandatory to safeguard equipment warranty by the manufacturer. The use of incompatible disinfectants or those not recommended renders the manufacturer warranty (and potentially the service contract) invalid, irrespective of any perceived damage, or lack thereof, to probes. Secondly, the use of a specific disinfectant may be recommended by a manufacturer, even though that disinfectant may not be available within the UK. For example, glutaraldehyde is commonly used in the USA and may be recommended for a probe but it is not available in the UK and the use exposes a department to legal action through the

HSAWA and COSHH. Guidance listing the information to be supplied by manufacturers of probes and disinfectants can be sought by referencing the Medical Devices Agency.¹⁰

This document cannot be exhaustive in giving trade names of detergents or disinfectants. It is suggested that you consult with your Infection Prevention team and/or endoscopy unit for advice on what is available for use in your establishment, e.g. type of nonlinting wipe with a neutral detergent for probe pre-cleaning.

Choice of disinfectant

A wide range of products exist (*Table 2*), but the choice of disinfectant should be governed by the microbicidal range, safety, and compatibility with the TOE probe.^{1,11} Agents used to date include aldehydes, hydrogen peroxide, peracetic acid, chlorine dioxide, superoxidized water, and alcohols. The use of alcohols and aldehydes as a disinfectant is discouraged owing to their fixative properties, resulting in protein (including prion protein) retention on the probe.¹

Transoesophageal echocardiography facility

Ideally, a suitable location (*Figure 3*) for the performance of TOE should comprise a two-room facility: one in which the procedure is performed and a separate room for decontamination.

 Table 2
 Comparison of disinfectants and their characteristics

Agent	Microbicidal efficacy (min)				Compatibility	Inactivation by
	Spores	Mycobacteria	Bacteria	Viruses		organic matter
Peracetic acid	5–10	5–10	<5	<5	Varies	No
Chlorine dioxide	5-10	5-10	<5	<5	Varies	Yes
Superoxidized water	10	<5	<5	<5	Varies	Yes

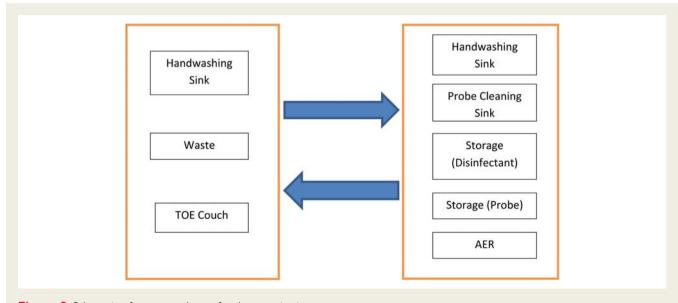


Figure 3 Schematic of two-room layout for decontamination.

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The procedure room should include space for hand washing, disposal of health-care waste (including sharps and non-sharps), and safe and secure TOE probe storage.

The decontamination room should include space for a sink for cleaning the probe, a separate sink for hand washing, disinfectant storage, and for the automated endoscope reprocessor (AER) if already in place, or facilities for manual disinfection instead. There should be a clear flow of work within this room with distinct and separate dirty and clean areas.

In facilities where the TOE procedure and disinfection are to take place in the same room, there should be pre-designated 'dirty' (pre-decontamination) and 'clean' (post-decontamination) areas to ensure that fully decontaminated probes cannot be confused with non- or partially decontaminated probes. Separating dirty and clean areas is a major step in eliminating probe recontamination or mistakenly using a probe that has not been fully decontaminated.

There should be a clearly identified location for storage of decontaminated probes. This must be situated within the 'clean' area of the room, so that there is no risk of recontamination of disinfected probes.

If two rooms are to be used, it is recommended that a rigid case should be available for transport of probes to minimize the risk of damage.

Storage of transoesophageal echocardiographic probes

TOE probes should not be stored in their delivery cases. This is a high-risk strategy—if one incompletely decontaminated probe is ever put in the case, the case will recontaminate all subsequent probes.

Ideal storage would be to hang the probe in a locked cupboard. An alternative would be to store the probe in a rigid tray (1-2 days)—longer storage may result in distortion of the probe shaft. Trays do not need to be sterile but should be visibly clean. The use of a tray liner and cover system may be beneficial particularly in transporting the probes, for example, using a green = clean and red = contaminated cover. This is widely used for flexible endoscopes and would be useful.

There is no time limit to storage of a clean probe in a clean area, as recommended above. Minimal environmental (i.e. non-patient derived) contamination should not pose any risk to patients in this context. There are no lumens within which microorganisms can grow. There is no need to reprocess if stored in a closed cupboard or a covered tray before patient use.

Protective sheaths

Sheaths are additional physical barriers to infection and probe damage. They do not, however, cover the whole of a TOE and so do not remove the possibility of cross-infection from the material deposited on the probe handle, cable, or plug socket (*Figure 2*). Sheaths are subject to perforation which may be undetectable to the naked eye. Perforation rates as high as 4.4% have been reported and may require a post-use air tightness test to confirm maintenance of structural integrity. Therefore, TOE probes should undergo the same decontamination whether or not a sheath is used.

Technique and facilities for decontamination

The following is a step-through guide to the guidelines for decontamination.

Pre-assessment

Standard infection control precautions should be adopted for each patient, irrespective of the status. However, placing of patients deemed to be of higher risk of carriage of infectious microorganisms towards the end of a list may allow more time for disinfection of relevant surfaces within the room.

If a sheath is to be used during TOE, prior latex allergy or intolerance should be established. Alternative sheaths¹⁴ (e.g. polyurethane) should be trialled in positive cases since potentially life-threatening reactions have previously been reported^{15,16} or the procedure could be done without a sheath.

Transoesophageal echocardiography facility

Before and after each patient, all probe and hand contact surfaces should be thoroughly wiped with a disinfectant.

Personnel

Before and after each patient, thorough hand washing should be carried out by all those present. The hands are the most common source of spreading infection.¹⁷ The majority of microorganisms may be removed by a short and thorough washing of the hands.¹⁸

Personnel involved in the TOE procedure should wear singleuse gloves and an apron. If splashing of bodily fluid is likely, then face and eye protection should be considered.

Probe cleaning

Decontamination should commence immediately following probe removal from the patient. If a sheath is used, this should be removed directly.

The probe shaft and probe tip should then be wiped with a single-use non-linting wipe moistened with detergent solution to remove gross contamination. It is suggested that you consult with your Infection Prevention team and/or endoscopy unit for advice on what is available for use in your establishment, e.g. type of non-linting wipe with neutral detergent.

A separate, similar second wipe (the first wipe will now be too contaminated to clean effectively) should then be used to wipe other parts of the probe to include non-immersible parts higher up, including the handle and plug socket. This second wipe reduces the potential for contamination on the housing unit, etc. from the operator's hand.

The TOE probe should then be carefully inspected to ensure that there has been no structural damage to cause loss of structural integrity of the coating that could allow ingress of contaminated material. Leak testing to demonstrate holes in the probe (different from electrical leak testing), as performed with gastroscopes, cannot be carried out with TOE probes as there is no port to permit insufflation of air.

The TOE plug socket should then be disconnected from the machine and the probe transported to the pre-designated decontamination area in a covered rigid container.

If the initial wipe has not thoroughly removed all visible contamination, the TOE probe should without delay then be immersed (while sparing the housing unit) in a sink or wash basin utilizing a detergent made up to the dilution and contact times stipulated by the manufacturer. It is essential for effective disinfection that any organic matter that may inhibit disinfectant action and shield microbes is removed. Immersion of the probe in a detergent solution at a specified concentration for a specified time as a standard procedure would represent best practice. If necessary, the probe should be wiped with a cloth under the water level to remove all remaining visible contamination.

Following detergent use, the probe should be thoroughly rinsed with potable-quality water to remove any residual detergent—if the detergent is not compatible with the disinfectant used in the next step.

Probe disinfection

Probe cleaning must be followed by probe disinfection between every patient. Probe disinfection may be automated or manual. Automated disinfection has the highest quality assurance for TOE probe decontamination and should be considered best practice. Manual disinfection is an alternative while working towards best practice.

Manual disinfection

Methods include the use of disinfectant wipes and baths. If manual disinfection is to be performed, particular care must be taken to ensure that disinfection is carried out not only to the probe tip and shaft but also to the handle, cable, and sections of the socket. It is important to ensure strict adherence to the manufacturer's instructions. Steps to be taken are as follows:

- (i) remove a wipe from a closed sachet;
- (ii) unfold the wipe and lay out on the palm the operator's hand;
- (iii) cover the wipe with disinfectant solution to the volume recommended by the manufacturer, ensuring that there is no delay between dispensing and use;
- (iv) wipe the whole TOE surface until it has been covered with disinfectant;
- (v) all areas of the surface must come into contact with the wipe at least once for the recommended contact time;
- (vi) discard the wipe to clinical waste.

Rinse thoroughly after disinfection to remove disinfectant residues after processing.

Automated cleaning

Some AERs will accommodate TOE probes by allowing immersion of the probe shaft in fluids and protecting the probe handle and socket from fluid exposure. This will provide a standardized decontamination procedure. However, the non-exposed parts of the probe will require manual decontamination as described above.

Detergents to be used within AERs should again be in accordance with manufacturer guidelines. Following AER use and as part of the complete AER cycle, further rinsing with sterile water and drying in air is recommended.

The use of dedicated AERs for automated decontamination of endoscopes is validated and recommended for use if available. ^{2,19} The advantages of this method are efficacy, reproducibility, and

diminished staff and patient exposure to potentially toxic disinfectants. The disadvantages are the high capital and running costs. The AER unit itself may become a source of contamination⁷ and therefore needs to be subjected to a self-disinfection procedure at the start of each working day.

Successful modification (development of sealant around the housing apparatus) of AERs to enable total TOE probe immersion has been reported recently. The study numbers are too small at this stage to recommend routine longer term use.²⁰

All AERs should be validated and tested in accordance with up to date NHS Estates Health Technical Memorandum (HTM) 2030 guidance¹⁹ (to be replaced by HTM 01-06 in the near future).

Rinse water should be potable quality if manual cleaning/disinfection/rinsing is used or, if an AER is used, it should be within the limits specified by HTM 2030 [or Choice Framework for local Policy and Procedure (CFPP) 01-06 when published].

Staff training, education, and quality assurance

Although different from GI endoscopy, the echocardiographer and allied health-care professionals should be aware of the responsibilities outlined in the Health Act 2006.²¹ This act emphasizes the need for staff involved with decontamination to:

- be trained and hold appropriate competences for their role;
- have monitoring systems to ensure that processes are fit for purpose;
- have tracking systems in place to ensure quality;
- identify and track patients after device usage in the event of future complication/infection.

Following an incident that resulted in inadequate decontamination, MHRA guidance (2004/28)³ review of practice highlighted the following as particular problems to avoid, including: incompatibilities between endoscope and AERs; staff being unfamiliar with decontamination processes; and poor communication between manufacturers of endoscopes and AERs. In order to specifically take these above points into account, it is recommended that:

- (i) When purchasing a TOE, an AER, or disinfectants, the manufacturer/supplier should be contacted to ensure compatibility and provide training tools.
- (ii) Regular audit of TOE decontamination practices is undertaken which may form the basis of a Care Quality Commission external examination.
- (iii) Each trust should have a decontamination network in place to include a nominated lead, operational manager, and operator/user.
- (iv) The User's responsibilities should include: certification of decontamination equipment as being fit to use, overseeing maintenance work to include quality assurance, and training other operators to perform similar tasks.
- (v) In the event of adverse incidents or concerns related to equipment failure/disinfectant malfunction, the MHRA should be contacted.
- (vi) A blame-free culture of incident reporting should be encouraged.

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Record keeping

Each probe should have a unique identifier and a record of the probe used on each patient and the decontamination procedure should be retained in the patient records and/or the unit records.

Special considerations

Prion infection

Providing decontamination of the TOE is to approved standards and the use of the instrument is deemed to be of low risk in relation to transmission of prion infection. Advice relating to prion infection is updated regularly and attention should be drawn to the following sites ^{22,23} for guidance:

http://www.dh.gov.uk/ab/ACDP/TSEguidance/index.htm#jumpTo3 and http://www.dh.gov.uk/prod_consum_dh/groups/dh_digital assets/@dh/@ab/documents/digitalasset/dh_117682.pdf

Conclusions

This document proposes a working solution to disinfection that can be implemented in Cardiology Departments for safe practice in TOE. There is little evidence relating to infection control within TOE, so the information provided represents a consensus of opinion. There are effectively two procedures possible within this document, a wipe-based disinfection system and a process based on automated processor disinfection. The former can be achieved with little additional modification of the current departments but the latter will require additional investment in many. Correct application of either of these processes is important not only for patient care but also to ensure that it is possible to prove that proper decontamination has taken place and that a given TOE procedure is not at fault should a patient be subsequently found to have blood-borne viral infection.

Summary of key recommendations for decontamination

1. Health and safety at work

Ensure compliance with HSAWA.

Follow guidance from the COSHH.

Departmental policies should be in place in the event of disinfectant spillage or staff sensitivity to a disinfectant.

Wear single-use gloves, goggles, and aprons when handling disinfectants.

2. Safeguarding manufacturer warranty

Check cross-compatibility of disinfectants and AERs. Reference MDA Device Bulletin.

3. Disinfectants

Check disinfectant compatibility with TOE probes and AERs.

Check disinfectant concentrations and contact times as per the manufacturers' recommendations.

Glutaraldehyde-based disinfectants should not be used in the UK. Alcohol-based products should not be used for disinfecting TOE probes.

4. Protective sheaths

Sheaths may reduce the level of probe contamination but their contribution to infection prevention does not reduce the need for further probe decontamination.

Consider polyurethane types in the event of latex intolerance/ allergy.

Only an air tightness test confirms non-perforation.

Decontamination technique should remain the same as for sheathless procedures and if used, decontamination must continue to be performed to cover transmission of infection from the probe handle, cable, and socket.

5. Workplace

Ensure a nearby sink or wash basin for cleaning and rinsing. Pre-designated 'dirty' and 'clean' areas should be in place.

Restrict entry to staff familiar with decontamination practices.

Probes should not be stored in a locked supposed on in a sigid to:

Probes should be stored in a locked cupboard or in a rigid tray. Consider transportation of probes using a tray liner and cover system.

6. Decontamination technique

(a) (i) Manual

Commence cleaning immediately following probe withdrawal. Initial wiping, followed by rinsing and near maximal immersion in disinfectant (sparing the probe housing unit).

Disinfectant to be made up to manufacturers' recommended concentrates and contact times.

And/or

(ii) Automated

Only use AERs if compatible and recommended by manufacturers. If the AER use does not comply with manufacturers' recommendations, it is possible that warranty and service contracts will be invalidated.

Disinfectant to be made up to the disinfectant manufacturers' recommended concentrates and contact times.

Rinsing with sterile water and air dying following AER use should be considered part of the AER cycle.

- (b) Manual disinfection of those parts of the probe not immersible in the AER.
- (c) Wipe all work surfaces clean.
- (d) Hand washing.

7. Automated endoscope reprocessors

Routine AER self-disinfection.

Culture rinse water weekly.

Change water filters as per the manufacturer guidance.

8. Staff training

Register with the Care Quality Commission.

Facilitate equipment training or training opportunities with manufacturers.

9. Quality assurance

Designated network to include lead, operational manager, and operators/users.

Departmental Auditing.

10. Incident reporting and traceability

Blame-free incident reporting culture is encouraged.

Contact MHRA in event of suspected equipment/disinfectant malfunction.

A log of procedures and an identifiable probe label should be cross-referenced for recording and traceability.

Conflict of interest: none declared.

Appendix

Glossary of terms

Automated Endoscope Reprocessor. Washer-disinfector machine capable of disinfection and rinsing to a reproducible standard and where the performance can be validated and verified.

Cleaning. The physical removal of infectious agents (but not necessarily their destruction) and the organic material which can shield them from disinfectants.

Detergent. Chemical that suspends organic material making subsequent removal easier.

Disinfection. The process of reduction in viable infectious agents to a safe level.

Decontamination. The process of cleaning combined with disinfection or sterilization that makes medical devices safe for reuse.

Decontamination Lead. Person responsible for implementing the operational policy for decontamination within a health-care establishment.

Decontamination User. Person designated by management to take overall responsibility for the management of the decontamination equipment.

Decontamination Operator. Person with the authority to operate decontamination equipment.

Sterilization. The process of rendering objects free from all viable microorganisms.

Areas of exclusivity: Clean area, the area post-decontamination; dirty area, the area pre-decontamination.

Components of the transoesophageal probe: Probe shaft, flexible with identifiable markers to assess the depth of insertion; probe

handle, contains both wheel and lock devices with steering buttons; probe tip, the flexible scan head housing the transducer, usually7.5 mm width \times 5.5 mm height \times 18.5 mm length, although new probe tips may be smaller; plug socket, end of the cable to attach the probe to its associated equipment.

References

- http://www.bsg.org.uk/clinical-guidelines/endoscopy/guidelines-fordecontamination-of-equipment-for-gastrointestinal-endoscopy.html.
- Department of Health. Health Technical Memorandum 01-01: Decontamination of Reusable Medical Devices, Part A. London: HSMO; 2007.
- 3. MHRA Medical Device Alert 2004/028.
- 4. http://www.4sonora.com/news/press/TEE_Probe_Care_Rev_3a1c.pdf.
- Sprach D, Silverstein FE, Stamm WE. Transmission of infection by gastrointestinal endoscopy and bronchoscopy. Ann Int Med 1993;118:117–28.
- Rutala WA, Weber DJ. Disinfection of endoscopes: review of new chemical sterilants used for high-level disinfection. Infect Control Hosp Epidemiol 1999;20:69–76.
- Levy P-Y, Teysseire N, Etienne J, Raoult D. A nosocomial outbreak of Legionella pneumophilia caused by contaminated transoesophageal echocardiography probes. Infect Control Hosp Epidemiol 2003;24:619–22.
- 8. Health and Safety Executive. Health and Safety Regulation: A Short Guide. London: HSE; 2003. http://hse.gov.uk/pubns/hsc13.pdf.
- Health and Safety Executive. COSHH: A Brief Guide to the Regulations: What You Need to Know About the Control of Substances Hazardous to Health 2002 (COSHH) Regulations. London: HSE; 2005. www.hse.gov.uk/pubns/indg136.pdf.
- Medical Devices Agency. Device Bulletin (DB) 2006 (05) Managing Medical Devices. Guidance for Healthcare and Social Services Organisations. London: MDA; 2006. www.mhra.gov.uk/publications/Safetyguidance/DeviceBulletins/CON2025142.
- 11. Niven K. An evaluation of chemical disinfecting agents used in endoscopy suites in the NHS. Health and Safety Executive. Research Report 445. 2006.
- Cote G, Denault A. Transoesophageal echocardiography-related complications. Can J Anesth 2008;55:622–47.
- Fritz S, Hust MH, Ochs C, Gratwohl I, Staiger M, Braun B. Use of latex cover sheath for transoesophageal echocardiography (TEE) instead of regular disinfection of the echoscope? Clin Cardiol 1993;16:737–40.
- Korsten HH, Peels CH, Heesakkers P, Rommelse K. Sheath placement over transoesophageal probes: a description of a 'self-moving' method. *Anesthesiology* 2002; 96:519–20
- Nawa Y, Imaizumi H, Masuda Y, Hazama K, Sato M, Namiki A. A case of anaphylaxic shock due to latex glove used on internal examination and on the probe of intrauterine echogram. Masui 2000;49:1027–9.
- Laxenaire MC. Drugs and other agents involved in anaphylactic shock occurring during anesthesia: a French multicentre epidemiological inquiry. Ann Fr Anaesth Reanim 1993;12:91–6.
- 17. Mortimer EA, Wolinsky E, Gonzaga AJ, Rammelkamp CH Jr. Role of airborne transmission in staphylococcal infections. *Br Med J* 1966;**1**:319–22.
- Sprunt K, Redman W, Leidy G. Antibacterial effectiveness of routine hand washing. *Pediatrics* 1973;52:264–71.
- Department of Health Estates and Facilities. Health Technical Memorandum 2030: Washer-Disinfectors: Validation and Verification. London: HSMO; 1997.
- Richardson A, Cowan C, Waters J, Diprose P. The development of a new technique for the decontamination of transoesophageal echocardiography probes. Infect Prev 2010;11:91–4.
- The Health Act 2006. Code of Practice for the Prevention and Control of Health Care
 Associated Infections. Department of Health. www.dh.gov.uk/assetRoot/04/13/93/
 37/04139337.pdf.
- 22. http://www.dh.gov.uk/ab/ACDP/TSEguidance/index.htm#jumpTo3.
- http://www.dh.gov.uk/prod_consum_dh/groups/dh_digitalassets/@dh/@ab/documents/digitalasset/dh_117682.pdf.